PROPORTION OF ROOT-DERIVED ACID PHOSPHOMONOESTERASE IN TOTAL SOIL ACID PHOSPHOMONOESTERASE IN DIFFERENT FORESTS

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Abstract

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Enzyme acid phosphomonoesterase (APM) plays an important role in phosphorus mineralization in different type of terrestrial ecosystems. This enzyme is of great agronomic significance because it hydrolyses organic phosphorus to different forms of inorganic phosphorus which are assimilable by plants. APM may also indicate changes in the quantity and quality of phosphorylated substrates in soil and is a good indicator of its biological state as well as presence of pollutants. APM may be produced by plant roots and soil microorganisms and both of these sources may play different role in phosphorus mineralization in different ecosystems. The aim of this work was determine acid phosphomonoesterase (APM) activity location in soil of different forest ecosystems. The APM activity location determination was performed on the basis of root-derived and soil-derived APM and expression of proportion of those root-derived in total soil APM up to 13 cm depth. The results of this preliminary study showed that root-derived APM formed 21–34% of total soil APM in pine and oak forest

acid phosphomonoesterase, soil, forests, roots

Extracellular phosphomonoesterases (orthophosphoric monoester phosphohydrolases) catalyze hydrolysis of a variety of organic phosphomonoesters and are therefore important in soil organic P mineralization and plant nutrition. Acid phosphomonoesterase (APM) in soil is located in different compartments. These include soil and its different fractions, plant roots and mycorrhizal fungi (Burns et al., 1972; McElhinney and Mitchell, 1993; Asmar and Gissel-Nielsen, 1997; Tamás et al., 2008). In these compartments, APM may occur intracellular or extracellular. As many studies were performed to determine localization of APM, still, the determination of proportion of root-derived APM in total soil environment of different terrestrial ecosystems is presently lacking. Due to this reason we have performed a pivotal experiment to determine proportion of root-derived APM in total soil APM of different forest ecosystems.

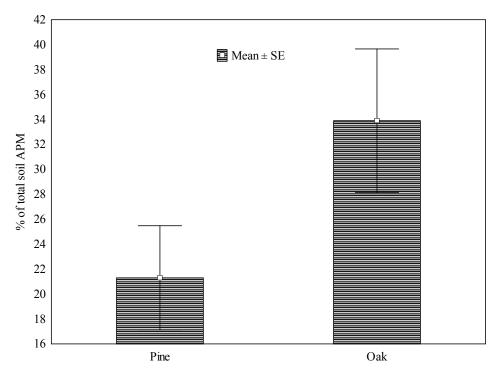
MATERIAL AND METHODS

The research was conducted in two forest plots. Oak forest and pine forest located at Training Forest Enterprise Masaryk Forest (TFE) Křtiny (oak forest N 49°16'51.656", E 16°37'55.646", pine forest N49°15'40.661", E 16°37'29.17"). Soil samples were taken at the beginning of December 2009 using PVC cylinders (height 15cm x inner diameter 5.9cm) which were inserted in pairs to a depth of 13 cm at five selected points of each of the localities (totally 10 samples per locality). After transportation to the laboratory in plastic bags, one of the pair of cylinders was carefully separated to organic and mineral layers whereas the other was used for separation of roots. The study supposed equivalence of each of cylinders in the pair. APM of soil as well as roots was determined using modified method of Rejšek (1991), and recalculated per total amount of soil and roots.

RESULTS AND DISCUSSION

The results of this primary study showed that APM derived from roots, representing rootassociated or released as a part of root exudates formed 21.3% in pine forest and 33.9% in oak forest of total soil APM up to depth of 13 cm (Fig. I). No statistically significant (P > 0.05) differences were found between the studied forests. These data were obtained at the beginning of December and no comparable study was performed so far on any type of ecosystems. APM in root exudates of different plants was found to form smaller fraction of root - derived APM ranging from 1.5 to less than 10% (Gaume et al., 2001; Asmar and Gissel-Nielsen, 1997; George et al., 2008). Phosphatase location seems to be uneven within root systems. Fine roots of Norway spruce were found to have significantly higher APM than medium or coarse roots (Firsching and Claassen, 1996). This aspect

was not evaluated in our study. Asmar and Gissel-Nielsen (1997) reported that in barley, most of the extracellular root APM activity to be associated with roots, that is 89.5-97.5%, while 1-5% was associated with rhizoplane microorganisms. Concerning soil APM, the highest activity was reported in the upper humus layer decreasing with soil depth (Taylor et al., 2002; Wittmann et al., 2004; Niemi et al., 2005.). APM in soil may also be influenced by plant coverage and its density due to support of microbial activity via plant roots (Dinesh et al., 2004). Higher APM activity was reported in rhizosphere compared to bulk soil (Tarafdar and Claassen, 1988; George et al., 2002; Hernesmaa et al., 2005; Zhao et al., 2007 etc.). As no comparable data related to this study are available for any type of ecosystem further more detailed study is necessary to evaluate the role of root derived APM in total soil AMP of different terrestrial ecosystems.



1: Proportion of root-serived APM on total soil APM in different type of forests

CONCLUSION

This opening study on location of acid phosphomonoesterase in soil showed dominance of soil-derived acid phosphomonoesterase activity up to 13 cm in pine and oak forests. The proportion of actually root-derived acid phosphomonoesterase was only 21–34%. In terms of aiming the future investigations, evaluating the role of roots in phosphorus transformation in different terrestrial ecosystems appears to be particularly challenging.

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REFERENCES

- ASMAR,F.,GISSEL-NIELSEN,G.,1997:Extracellular phosphomono- and phosphodiesterase associated with and released by the roots of barley genotypes: a non-destructive method for the measurement of the extracellular enzymes of roots. *Biology and Fertility of Soils* 25, 2: 117–122. ISSN 0178-2762.
- BURNS, R. G., PUKITE, A. H., MCLAREN, A. D., 1972: Concerning the location and persistence of soil urease. *Soil Science Society of America Journal*, 36, 308–311. ISSN 0361-5995.
- DINESH, R., SURYANARAYANA, M. A., GHOSHAL CHAUDHURI, S., SHEEJA, T. E., 2004: Longterm influence of leguminous cover crops on the biochemical properties of a sandy clay loam Fluventic Sulfaquent in a humid tropical region in India. *Soil and Tillage Research* 77, 1: 69–77. ISSN 0167-1987.
- FIRSCHING, B. M., CLAASSEN, N., 1996: Root phosphatase activity and soil organic phosphorus utilization by Norway spruce [*Picea abies* (L.) Karst.]. *Soil Biology and Biochemistry* 28, 10–11: 1417–1424. ISSN 0038-0717.
- GAUME, A., MÄCHLER, F., DE LÉON, C., NARRO, L., FROSSARD, E., 2001: Low-P tolerance by maize (*Zea mays* L.) genotypes: Significance of root growth, and organic acids and acid phosphatase root exudation. *Plant and Soil* 228, 2: 253–264. ISSN 0032-079X.
- GEORGE, T. S., GREGORY, P. J., WOOD, M., READ, D., BURESH, R. J., 2002: Phosphatase activity and organic acids in the rhizosphere of potential agroforestry species and maize. *Soil Biology and Biochemistry* 34, 10: 1487–1494. ISSN 0038-0717.
- GEORGE, T. S., GREGORY, P. J., HOCKING, P., RICHARDSON, A. E., 2008: Variation in root-associated phosphatase activities in wheat contributes to the utilization of organic P substrates in vitro, but does not explain differences in the P-nutrition of plants when grown in soil. *Environmental* and *Experimental Botany* 64, 3: 239–249. ISSN 0098-8472.
- HERNESMAA, A., BJÖRKLÖF, K., KIIKKILÄ, O., FRITZE, H., HAAHTELA, K., ROMANTSCHUK, M., 2005: Structure and function of microbial communities in the rhizosphere of Scots pine after tree-felling. *Soil Biology and Biochemistry* 37, 4: 777–785. ISSN 0038-0717.

- McELHINNEY, C., MITCHELL, D. T., 1993: Phosphatase activity of four ectomycorrhizal fungi found in a Sitka spruce-Japanese larch plantation in Ireland. *Mycological Research* 97, 6: 725–732. ISSN 0953-7562.
- NIEMI, R. M., VEPSÄLÄINEN, M., WALLENIUS, K., SIMPANEN, S., ALAKUKKU, L., PIETOLA, L., 2005: Temporal and soil depth-related variation in soil enzyme activities and in root growth of red clover (*Trifolium pratense*) and timothy (*Phleum pratense*) in the field. *Applied Soil Ecology* 30, 1: 113–125. ISSN 0929-1393.
- REJŠEK, K., 1991: Acid phosphomonoesterase activity of ectomycorrhizal roots in Norway spruce pure stands exposed to pollution. *Soil Biology and Biochemistry* 23, 7: 667–671. ISSN 0038-0717.
- TAMÁS, L., DUDÍKOVÁ, J., ĎURČEKOVÁ, K., HUTTOVÁ, J., MISTRÍK, I., ZELINOVÁ, V., 2008: The impact of heavy metals on the activity of some enzymes along the barley root. *Environmental* and *Experimental Botany* 62, 1: 86–91. ISSN 0098-8472.
- TARAFDAR, J. C., CLAASSEN, N., 1988: Organic phosphorus compounds as a phosphorus source for higher plants through the activity of phosphatases produced by plant roots and microorganisms. *Biology and Fertility of Soils* 3, 4: 199–204. ISSN 0178-2762.
- TAYLOR, J. P., WILSON, B., MILLS, M. S., BURNS, R. G., 2002: Comparison of microbial numbers and enzymatic activities in surface soils and subsoils using various techniques. *Soil Biology and Biochemistry* 34, 3: 387–401. ISSN 0038-0717.
- WITTMANN, CH., KÄHKÖNEN, M. A., IL-VESNIEMI, H., KUROLA, J., SALKINOJA-SA-LONEN, M. S., 2004: Areal activities and stratification of hydrolytic enzymes involved in the biochemical cycles of carbon, nitrogen, sulphur and phosphorus in podsolized boreal forest soils. *Soil Biology and Biochemistry* 36, 3: 425–433. ISSN 0038-0717.
- ZHAO, Q., ZENG, D. H., LEE, D. K., HE, X. Y., FAN, Z. P., JIN, Y. H., 2007: Effect of *Pinus sylvestris* var. mongolica afforestation on soil phosphorus status of the Keerqin Sandy Land in China. *Journal of Arid Environments* 69, 4: 569–582. ISSN: 0140-1963.

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